



Steroidogenesis in luteal cell: A critical pathway for progesterone production

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ABSTRACT

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Received: September 27, 2014 Accepted: December 22, 2014 Published: January 20, 2015 The ovarian cycle is central to the physiology of reproduction in most domestic mammals; characterized by repeated patterns of cellular proliferation, differentiation and transformation of ovarian steroidogenic cells (granulosa, theca and luteal cells) that accompany follicular development, ovulation, followed by formation, function and regression of the corpus luteum (CL). Luteal cells are the chief cells present in the CL. Steroid hormones are synthesized in the adrenal gland, gonads, and placenta critical for normal reproductive function. Steroidogenesis is the main process that occurs in luteal cells to produce the progesterone that is essential for the establishment and maintenance of pregnancy. Steroidogenase enzyme are major regulators of steroidogenesis process in the luteal cells. Progesterone regulates the length of the estrus cycle by influencing the timing of the luteolytic PGF2 α signal from the endometrium. Termination of estrus cycle is characterized by functional luteolysis and structural luteolysis. Functional luteolysis is due to regression of CL leading to loss of steroidogenic capacity. Finally, luteal cell death and resorption makes the way to structural luteolysis. Therefore, it is paramount to know the exact mechanism of progesterone production through steroidogenesis process in luteal cells. This review is to highlight the important steps of steroidogenesis process in luteal cells.

KEY WORDS: Luteal cell, steroidogenesis, progesterone, steroidogenic acute regulatory protein, cytochrome P450 side chain cleavage enzyme, 3β hydroxylsteroid dehydrogenase enzyme

INTRODUCTION

Luteal cells are the structural and functional units of mammalian corpus luteum (CL). Formation of the CL is initiated by series of morphologic and biochemical changes in theca interna (TI) and granulosa cells (GC) of pre-ovulatory follicle (POF). These changes are termed as luteinization, occurs after the preovulatory luteinizing hormone (LH) surge. The mammalian CL is a heterogeneous tissue composed of steroidogenic, small luteal cells (SLCs) and large LCs (LLCs), and non-steroidogenic cells, such as fibroblasts, endothelial and immune cells [1-3]. The CL is a transient endocrine organ with an intense angiogenesis, secretes a large amount of progesterone (P4), required for establishment and maintenance of pregnancy [4,5]. Steroidogenesis mainly occurs in the luteal cells of CL. SLCs are more responsive to LH but LLCs mainly responsible for P4 secretion. Key P4 secretion factors are steroidogenic acute regulatory protein (StAR), cytochrome P450 side chain cleavage enzyme (Cyt. P450 scc) and 3 beta hyroxysteroid dehydrogenase enzyme $(3\beta$ -HSD) [6-8]. Unlike most of the adult tissues, CL exhibits regular periods of growth through angiogenesis, functions by production of P4 and finally regresses. If the pregnancy does not occur, the CL has to regress to commence the next reproductive cycle again. Optimum level of P4 is

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critical for the development of endometrium and implantation. Therefore, insufficient P4 will cause early embryonic mortality leading to infertility. Foley, 1996 [9] reported that reduced P4 secretion due to luteal dysfunction resulted in infertility of domestic animals. Luteal insufficiency is considered as one of the primary causes of embryonic mortality in Buffalo [10]. Therefore, a good understanding of steroidogenesis mechanism would improve the reproductive efficiency and productivity of domestic livestocks.'

OVARIAN LUTEAL DYNAMICS

Follicular - luteal transition is a dynamic process, which involves a series of biochemical and morphological changes in the POF following LH surge [11,12]. The preovulatory LH surge results in luteinization of granulosal (converted to LLCs) and thecal cells (converted to SLCs) and alters the steroidogenic pathway so that P4, the primary steroid hormone is produced by these luteal cells [13]. In ewes, SLCs (12-20 μ m in diameter), thought to be of follicular thecal cell origin, contain receptors for LH, with a 5-15 fold increase in secretion of P4 via activation of the cyclic adenosine monophosphate (cAMP)/(protein kinase A) PKA second messenger pathway [11]. LLCs (>20 μ m) are primarily of GC origin, secrete high basal concentrations of P4 (> 85%), although LH receptors are present, do not respond to LH or cAMP the mechanism of which is still unknown [14,15]. PGF2 α causes luteolysis of LLCs by activating two second messenger pathways [15]. Activated protein kinase C directly inhibits progesterone synthesis leading to luteolysis, whereas inositol 1, 4, 5 tri phosphate (IP3) - Ca⁺² pathway increases of intracellular calcium concentrations resulting in luteal cell apoptosis [11,16].

LUTEAL CELL STEROIDOGENESIS

Steroidogenic Substrates

The substrate for steroidogenesis is cholesterol. Cholesterol is synthesized by liver and transported to steroidogenic tissues such as the adrenal cortex and gonads, in the form of lipoproteins [11,17]. The uptake of low-density lipoprotein by luteal cells occurs by receptor-mediated endocytosis (RME) [18] but Uptake of HDL occurs through an unknown mechanism, which is RME independent [19]. The fate of cholesterol inside cell is either to be stored in the form of cholesterol esters by cholesterol ester synthetase or to be used for steroidogenesis in mitochondria and smooth endoplasmic reticulum (SER) [11,15]. After depletion of free cholesterol, cholesterol esters are utilized steroidogenesis after hydrolysis by cholesterol esterase. Decreased lipoprotein concentration during lipid deprived conditions stimulates luteal cells to follow an alternate pathway to achieve the optimum level of cholesterol from acetate [15].

Cholesterol Transport

First step of P4 synthesis is the transport of cholesterol from the cytoplasm to the inner mitochondrial membrane, which is the rate-limiting step in hormone biosynthesis. This transport process is most acutely influenced by second messengers. Three essential proteins are involved in transport of cholesterol from the outer to the inner mitochondrial membrane. These three proteins are StAR, peripheral type benzodiazepine receptor (PBR) and endozepine, the natural ligand for PBR. StAR is synthesized as a 37 kDa protein that contains a mitochondrial targeting sequence [20,21] and multiple potential PKA and PKC phosphorylation sites [22]. PKA mediated phosphorylation of StAR stimulates cholesterol transport, whereas phosphorylation by PKC may inhibit this process. Sterol binding proteins also appear to play a role in the transport of cholesterol to the mitochondria [23]. StAR is the candidate protein, which transports the cholesterol to the inner membrane of mitochondria [8,24-27]. In bovine tissues, expression status of StAR m-RNA is very low in GC, moderate in early CL and maximum during midcycle, which is maintained throughout pregnancy [28]. StAR expression was limited to TI of POF and luteinized granulosa and theca cells of CL [29].

PBR present in the membranes of mitochondria in steroid producing cells, also appear to play a role in cholesterol transport from the outer to the inner mitochondrial membrane [30]. StAR binds to cholesterol in the cytosol and may directly transport it into inner mitochondrial membrane or may be through PBR [31]. Endozepine helps in cholesterol transport by changing the conformation of PBR enabling it to transport the cholesterol to the inner mitochondrial membrane or by facilitating the StAR to PBR interaction for exchange of cholesterol [32]. Phosphorylation of PBR by PKA enhances cholesterol transport [33].

Conversion of Cholesterol to Progesterone

Cytochrome P450 scc (an inner mitochondria membrane protein) catalyzes the conversion of cholesterol to the first steroid, pregnenolone [6,34]. Pregnenolone is then transported to the smooth endoplasmic reticulum, where 3β -HSD/ Δ^5 , Δ^4 isomerase converts pregnenolone to first biologically active hormone progesterone [7,35]. So for P4 production from luteal cells, free cholesterol needs to be translocated to the inner membrane of mitochondria to become available as a substrate at the site of P450scc [36]. During luteal development there is a gradual up-regulation in StAR as well as P450scc transcript levels, which reach a plateau at mid - luteal phase (8th to 12th days) in cattle [28]. In buffalo, P450scc, StAR and 3 β -HSD mRNA expression was high in mature CL and low during regressing stage [37].

CONCLUSION

Progesterone is the primary steroid hormone synthesized by luteal cells of CL. The main function of progesterone is to establish and maintain pregnancy in mammals. In most domestic species, early embryonic mortality is considered as one of the major causes of fertility loss. Therefore, impairment in the luteal steroidogenesis leads to the lower level of progesterone production that results in reduced reproductive efficiencies. All these above conditions finally end in major economic losses to Indian farmers. Thorough understanding in process of steroidogenesis and steps of progesterone synthesis from luteal cells will enable the researchers for making successful strategies to reduce the rate of early embryonic mortality; thereby enhance the pregnancy rates in domestic species, which could improve the socio economic status of farmers as well as the country.

REFERENCES

- 1. O'Shea JD, Rodgers RJ, D'Occhio MJ. Cellular composition of the cyclic corpus luteum of the cow. J Reprod Fertil 1989;85:483-7.
- Pate JL. Intercellular communication in the bovine corpus luteum. Theriogenology 1996;45:1381-97.
- Chouhan VS, Panda RP, Yadav VP, Babitha V, Khan FA, Das GK, *et al.* Expression and localization of vascular endothelial growth factor and its receptors in the corpus luteum during oestrous cycle in water buffaloes (*Bubalus bubalis*). Reprod Domest Anim 2013;48:810-8.
- Schams D, Berisha B. Regulation of corpus luteum function in cattle – an overview. Reprod Domest Anim 2004;39:241-51.
- Meidan R, Levy N, Kisliouk T, Podlovny L, Rusiansky M, Klipper E. The yin and yang of corpus luteum-derived endothelial cells: balancing life and death. Domest Anim Endocrinol 2005;29:318-28.
- Lambeth JD, Pember SO. Cytochrome P-450scc-adrenodoxin complex. Reduction properties of the substrate-associated cytochrome and relation of the reduction states of heme and iron-sulfur centers to association of the proteins. J Biol Chem 1983;258:5596-602.
- 7. Cherradi N, Rossier MF, Vallotton MB, Timberg R, Friedberg I, Orly J,

et al. Sub mitochondrial distribution of three key steroidogenic proteins (steroidogenic acute regulatory protein and cytochrome P450scc and 3b-hydroxysteroid dehydrogenase isomerase enzymes) upon stimulation by intracellular calcium in adrenal glomerulosa cells. J Biol Chem 1997;272:7899-7.

- Stocco C, Telleria C, Gibori G. The molecular control of corpus luteum formation, function, and regression. Endocr Rev 2007;28:117-49.
- Foley GL. Pathology of the corpus luteum of cows. Theriogenology 1996;45:1413-28.
- Campanile G, Neglia G. Embryonic mortality in buffalo cows. Ital J Anim Sci 2007;6 Suppl 2:119-29.
- Niswender GD, Juengel JL, Silva PJ, Rollyson MK, McIntush EW. Mechanisms controlling the function and life span of the corpus luteum. Physiol Rev 2000;80:1-29.
- 12. Reynolds LP, Redmer DA. Growth and development of the corpus luteum. J Reprod Fertil Suppl 1999;54:181-91.
- 13. Niswender GD, Nett TM. Corpus luteum and its control in infraprimate species. Physiol Reprod 1994;1:781-816.
- Niswender GD, Schwall RH, Fitz TA, Farin CE, Sawyer HR. Regulation of luteal function in domestic ruminants: new concepts. Recent Prog Horm Res 1985;41:101-51.
- 15. Niswender GD. Molecular control of luteal secretion of progesterone. Reproduction 2002;123:333-9.
- Wiltbank MC, Diskin MG, Niswender GD. Differential actions of second messenger systems in the corpus luteum. J Reprod Fertil Suppl 1991;43:65-75.
- 17. Krisans SK. Cell compartmentalization of cholesterol biosynthesis. Ann N Y Acad Sci 1996;804:142-64.
- Brown MS, Goldstein JL. A receptor-mediated pathway for cholesterol homeostasis. Science 1986;232:34-47.
- Lestavel S, Fruchart JC. Lipoprotein receptors. Cell Mol Biol (Noisyle-grand) 1994;40:461-81.
- Stocco DM, Sodeman TC. The 30-kDa mitochondrial proteins induced by hormone stimulation in MA-10 mouse Leydig tumor cells are processed from larger precursors. J Biol Chem 1991;266:19731-8.
- Clark BJ, Wells J, King SR, Stocco DM. The purification, cloning, and expression of a novel luteinizing hormone-induced mitochondrial protein in MA-10 mouse Leydig tumor cells. Characterization of the steroidogenic acute regulatory protein (StAR). J Biol Chem 1994;269:28314-22.
- Juengel JL, Niswender GD. Molecular regulation of luteal progesterone synthesis in domestic ruminants. J Reprod Fertil Suppl 1999;54:193-205.
- Ikonen E. Molecular mechanisms of intracellular cholesterol transport. Curr Opin Lipidol 1997;8:60-4.
- Wiltbank MC, Belfiore CJ, Niswender GD. Steroidogenic enzyme activity after acute activation of protein kinase (PK) A and PKC in ovine small and large luteal cells. Mol Cell Endocrinol 1993;97:1-7.
- 25. Stevens VL, Xu T, Lambeth JD. Cholesterol trafficking in

steroidogenic cells. Reversible cycloheximide-dependent accumulation of cholesterol in a pre-steroidogenic pool. Eur J Biochem 1993;216:557-63.

- Belfiore CJ, Hawkins DE, Wiltbank MC, Niswender GD. Regulation of cytochrome P450scc synthesis and activity in the ovine corpus luteum. J Steroid Biochem Mol Biol 1994;51:283-90.
- Wang X, Liu Z, Eimerl S, Timberg R, Weiss AM, Orly J, *et al.* Effect of truncated forms of the steroidogenic acute regulatory protein on intramitochondrial cholesterol transfer. Endocrinology 1998;139:3903-12.
- Hartung S, Rust W, Balvers M, Ivell R. Molecular cloning and *in vivo* expression of the bovine steroidogenic acute regulatory protein. Biochem Biophys Res Commun 1995;215:646-53.
- Kiriakidou M, McAllister JM, Sugawara T, Strauss JF 3rd. Expression of steroidogenic acute regulatory protein (StAR) in the human ovary. J Clin Endocrinol Metab 1996;81:4122-8.
- 30. Papadopoulos V, Brown AS. Role of the peripheral-type benzodiazepine receptor and the polypeptide diazepam binding inhibitor in steroidogenesis. J Steroid Biochem Mol Biol 1995;53:103-10.
- Kallen CB, Billheimer JT, Summers SA, Stayrook SE, Lewis M, Strauss JF 3rd. Steroidogenic acute regulatory protein (StAR) is a sterol transfer protein. J Biol Chem 1998;273:26285-8.
- Li H, Papadopoulos V. Peripheral-type benzodiazepine receptor function in cholesterol transport. Identification of a putative cholesterol recognition/interaction amino acid sequence and consensus pattern. Endocrinology 1998;139:4991-7.
- Papadopoulos V, Amri H, Boujrad N, Cascio C, Culty M, Garnier M, et al. Peripheral benzodiazepine receptor in cholesterol transport and steroidogenesis. Steroids 1997;62:21-8.
- Miller WL. Molecular biology of steroid hormone synthesis. Endocr Rev 1988;9:295-318.
- Hanukoglu I. Steroidogenic enzymes. Structure, function, and role in regulation of steroid hormone biosynthesis. J Steroid Biochem 1992;43:779-804.
- Privalle CT, Crivello JF, Jefcoate CR. Regulation of intramitochondrial cholesterol transfer to side-chain cleavage cytochrome P-450 in rat adrenal gland. Proc Natl Acad Sci U S A 1983;80:702-6.
- Kumar L, Panda RP, Hyder I, Yadav VP, Sastry KV, Sharma GT, et al. Expression of leptin and its receptor in corpus luteum during estrous cycle in buffalo (*Bubalus bubalis*). Anim Reprod Sci 2012;135:8-17.

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Source of Support: Nil, Conflict of Interest: None declared.